



#### **REVIEW**

# Recent neuroimaging, neurophysiological, and neuropathological advances for the understanding of NPC [version 1; referees: 3 approved]

Alberto Benussi<sup>1</sup>, Maria Sofia Cotelli <sup>10</sup>, Alessandro Padovani<sup>1</sup>, Barbara Borroni<sup>1</sup>

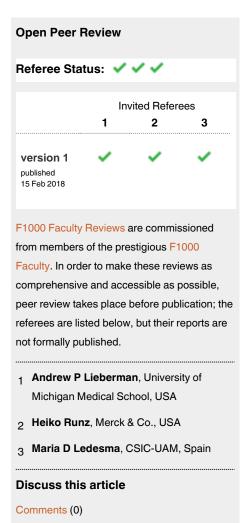
v1

**First published:** 15 Feb 2018, **7**(F1000 Faculty Rev):194 (doi: 10.12688/f1000research.12361.1)

Latest published: 15 Feb 2018, **7**(F1000 Faculty Rev):194 (doi: 10.12688/f1000research.12361.1)

#### **Abstract**

Niemann–Pick disease type C (NPC) is a rare autosomal recessive lysosomal storage disorder with extensive biological, molecular, and clinical heterogeneity. Recently, numerous studies have tried to shed light on the pathophysiology of the disease, highlighting possible disease pathways common to other neurodegenerative disorders, such as Alzheimer's disease and frontotemporal dementia, and identifying possible candidate biomarkers for disease staging and response to treatment. Miglustat, which reversibly inhibits glycosphingolipid synthesis, has been licensed in the European Union and elsewhere for the treatment of NPC in both children and adults. A number of ongoing clinical trials might hold promise for the development of new treatments for NPC. The objective of the present work is to review and evaluate recent literature data in order to highlight the latest neuroimaging, neurophysiological, and neuropathological advances for the understanding of NPC pathophysiology. Furthermore, ongoing developments in disease-modifying treatments will be briefly discussed.



<sup>&</sup>lt;sup>1</sup>Neurology Unit, Department of Clinical and Experimental Sciences, University of Brescia, Viale Europa, 11, 25123 Brescia BS, Italy <sup>2</sup>Neurology Unit, Valle Camonica Hospital, 25040 Esine, Brescia, Italy



Corresponding author: Barbara Borroni (barbara.borroni@unibs.it)

**Author roles: Benussi A**: Conceptualization, Writing – Original Draft Preparation; **Cotelli MS**: Data Curation, Writing – Review & Editing; **Padovani A**: Data Curation, Writing – Review & Editing; **Borroni B**: Conceptualization, Data Curation, Supervision, Writing – Original Draft Preparation

Competing interests: No competing interests were disclosed.

How to cite this article: Benussi A, Cotelli MS, Padovani A and Borroni B. Recent neuroimaging, neurophysiological, and neuropathological advances for the understanding of NPC [version 1; referees: 3 approved] F1000Research 2018, 7(F1000 Faculty Rev):194 (doi: 10.12688/f1000research.12361.1)

Copyright: © 2018 Benussi A et al. This is an open access article distributed under the terms of the Creative Commons Attribution Licence, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Grant information: The author(s) declared that no grants were involved in supporting this work.

First published: 15 Feb 2018, 7(F1000 Faculty Rev):194 (doi: 10.12688/f1000research.12361.1)

#### Introduction

Niemann–Pick disease type C (NPC) (MIM 257220) is an autosomal recessive neurovisceral lysosomal lipid storage disorder due to mutations of either the *NPC1* (95% of families)<sup>1</sup> or the *NPC2* gene<sup>2</sup>. The incidence of NPC is estimated to be 1 in 100,000 live births<sup>3–5</sup>, although the late-onset phenotypes or variant forms with visceral-only or neurologically mild presentations might have a much higher frequency<sup>6</sup>.

Although the exact functions of the *NPC1* and *NPC2* genes are still to be fully elucidated, mutations in these genes cause a loss of function, resulting in the accumulation of unesterified cholesterol and glycosphingolipids within the late endosome/lysosome of all cells, leading to downstream effects on cholesterol homeostasis<sup>7–9</sup>. Recently, using structural biology approaches, such as crystallography and cryo–electron microscopy, researchers have gained insights into how NPC1 and NPC2 proteins function in tandem to facilitate cholesterol efflux from late endosomes/lysosomes<sup>10–19</sup>.

The broad clinical spectrum ranges from a neonatal, rapidly fatal disease to an adult-onset chronic neurodegenerative disorder, and patients can be classified into four general categories based on age at neurological onset: the early infantile, late infantile, juvenile, and adolescent/adult onset form<sup>3</sup>. In most patients, disease severity is defined by the neurological involvement but usually is preceded by systemic signs such as neonatal cholestatic jaundice or isolated spleen or liver enlargement in infancy or childhood<sup>4</sup>. In the majority of patients with NPC, the liver disease frequently resolves with time<sup>3,4</sup>.

Clinical pictures of NPC are widely heterogeneous, and there are a number of neurological signs and symptoms with different ages at onset and different rates of progression: gait ataxia, clumsiness, cataplexy, epilepsy, dystonia, supranuclear gaze palsy, dysarthria, dysphagia, cerebellar ataxia, psychiatric illnesses, or cognitive decline<sup>3,4,20</sup>. Psychiatric disturbances may be undiagnosed for several years and are often characterized by psychosis such as paranoid delusions, visual or auditory hallucinations, or behavioral abnormalities<sup>20</sup>. The age of onset of neurological manifestations usually correlates with the patient's prognosis<sup>4</sup>.

Slightly different phenotypes have been observed in patients with *NPC2* gene mutations, which are frequently associated with a severe phenotype, characterized by pulmonary infiltrates, respiratory failure, and death in early age<sup>21–23</sup>. Adult-onset disease with frontal lobe atrophy has also been described<sup>24</sup>, and in some cases prolonged survival into middle adult life has been reported<sup>25</sup>.

Various NPC disability scales have been developed<sup>26–30</sup>; however, the original scale by Iturriaga,<sup>29</sup> and the subsequent modified versions are the most widely adopted<sup>27,28</sup>.

The diagnostic workup currently includes a combination of both biochemical and genetic analysis. Plasma cholesterol oxidation products (oxysterols), including cholestane- $3\beta$ , $5\alpha$ , $6\beta$ -triol<sup>31,32</sup> and filipin staining in cultured fibroblasts<sup>33–35</sup> are considered key in the diagnostic workup.

Plasma oxysterols have been shown to be highly sensitive for NPC; however, the originally reported specificity has recently been widened to other disorders of sterol metabolism, including acid sphingomyelinase deficiency (Niemann–Pick type A and B disease), cerebrotendinous xanthomatosis, and lysosomal acid lipase deficiency (Wolman disease)<sup>36–38</sup>. Furthermore, oxysterols can be increased in neonates with non-NPC cholestasis<sup>39</sup>.

Until recently, filipin staining was considered the gold standard assay for NPC diagnosis<sup>4,9,33,40</sup>, the typical pattern was observed in 80 to 85% of cases with NPC<sup>33,41–43</sup>, and a positive staining occurred in 80 to 100% of cells<sup>33</sup>. Only one condition, mucolipidosis II/III, is known to resemble a classic NPC filipin profile<sup>33</sup>. In the remaining 15 to 20% of NPC cases, a "variant" profile is observed, and only 50 to 80% cells stain positive<sup>33</sup>. A "variant" profile can also be observed in various conditions, including NPC heterozygous carriers<sup>44</sup>, Niemann–Pick type A and B disease<sup>45</sup>, MEGDEL (3-methylglutaconic aciduria, deafness, encephalopathy, and Leigh-like disease) syndrome<sup>46</sup>, Smith-Lemli-Opitz syndrome<sup>47</sup>, and Tangier disease<sup>48</sup>.

A major limitation of the filipin test is that it requires cell cultures of living fibroblasts from a skin biopsy, has relatively long turnaround times, and is performed in only a few specialized laboratories worldwide<sup>49</sup>.

Recently, a bile acid–based newborn screening for NPC, which identified  $3\beta,5\alpha,6\beta$ -trihydroxycholanic acid and its glycine conjugate, metabolites of cholestane- $3\beta,5\alpha,6\beta$ -triol, on dried blood spots, provided 100% specificity and sensitivity in identifying patients with NPC<sup>50</sup>.

Genetic testing, which involves the sequencing of the *NPC1* and *NPC2* genes, is also available; however, it is still inconclusive in 12 to 15% cases because of the unknown pathogenicity of the mutation, the lack of a study of allele segregation, and the possible existence of one unidentified mutant allele<sup>6</sup>.

A series of therapies for NPC are actively being developed. Miglustat, an iminosugar that is a competitive inhibitor of glucosylceramide synthase and specifically targets the metabolic pathway that leads to the synthesis of glycosphingolipids in neurons and other cells, has been approved for the treatment of NPC in Europe and elsewhere<sup>51</sup>.

Very recently, the most up-to-date clinical guidelines on NPC were published by an expert group<sup>52</sup>.

In recent years, numerous studies have increased the understanding of NPC, highlighting the very complex and multifaceted nature of the disease, and reported possible links to other neuro-degenerative disorders, such as Alzheimer's disease (AD) and frontotemporal dementia (FTD).

The objective of the present work is to review and evaluate recent literature data in order to highlight the latest neuroimaging, neurophysiological, and neuropathological advances for the understanding of NPC pathophysiology. Furthermore, recent developments in disease-modifying treatments will be briefly discussed.

#### Neuroimaging

Historically, brain magnetic resonance imaging (MRI) and computed tomography scans were not usually considered very useful for the diagnosis of NPC, as most of the findings are unspecific, sometimes showing cerebellar or cortical atrophy or, in the severe infantile form, white matter changes<sup>3</sup>. However, in recent years, numerous studies have shown the involvement of several cerebral structures in patients with NPC, even highlighting the possible modifications induced by therapeutic interventions (Table 1).

Voxel-based morphometry analyses have shown a significant involvement of the hippocampus, thalamus, striatum, and cerebellum in NPC<sup>53–57</sup>. In particular, cerebellar gray matter and left thalamus volume loss were significantly correlated with Iturriaga disability scale changes and ataxia measures<sup>53,58</sup>. Furthermore, untreated patients exhibited what may appear to be greater thalamic and cerebellar gray and white matter reductions over time compared with both controls and patients treated with miglustat<sup>53</sup>.

As in progressive supranuclear palsy, the pontine-to-midbrain ratio is increased in adult patients with NPC compared

with controls, and the strong correlation with illness and oculomotor variables suggests that it may be a useful marker for illness progression in NPC<sup>59</sup>.

The atrophy pattern in the thalamus, hippocampus, and caudate nucleus, observed with cortical thickness analyses, showed a significant correlation with memory, executive functions, and motor control dysfunction<sup>58</sup>.

The involvement of deep gray nuclei has also been confirmed by <sup>123</sup>I-FP-CIT (ioflupane I 123 DaTSCAN) single-photon emission computed tomography (SPECT) imaging in a case of NPC, showing a marked, symmetrical loss of dopamine transporter binding, especially in the putamen<sup>57</sup>. This pattern has also been observed in a heterozygote patient with a "variant phenotype" in filipin staining and with high levels of plasma oxysterols<sup>60</sup>.

Diffusion tensor imaging (DTI) analyses showed decreased fractional anisotropy in NPC patients compared with controls, especially in the corpus callosum, internal capsule, corona radiate, and the cingulate gyrus, with an early but transient improvement of DTI metrics after miglustat treatment<sup>61–63</sup>. Global callosal measures correlated significantly with duration of illness and symptom score and at trend level with degree of filipin staining<sup>64</sup>.

In agreement with these studies, myelin water imaging, a technique that measures the amount of water present within the myelin of white matter tracts<sup>65,35</sup>, has shown large reductions of myelin water fraction in large association tracts and the corpus

Table 1. Proposed diagnostic and prognostic markers for Niemann–Pick disease type C.

Variable	Marker	References
Brain MRI-VBM	Hippocampus, thalamus, striatum, cerebellum atrophy	53–57
Brain MRI	Pontine-to-midbrain ratio atrophy	59
Brain MRI-DTI	Corpus callosum, corona radiate, cingulate gyrus decreased fractional anisotropy	61–64
Brain H-MRSI	Frontal and parietal cortices, centrum semiovale, caudate nucleus decreased N-acetyl aspartate/ Creatine ratio	73,74
<sup>123</sup> I-FP-CIT (Ioflupane I 123 DaTScan)	Symmetrical loss of dopamine transporter binding	57–60
Brain FDG-PET	Frontal and temporal lobe hypometabolism	68,72
Brain SPECT	Frontal and temporal lobe hypoperfusion	69
TMS	SAI and LTP-like cortical plasticity impairment	60,93

DTI, diffusion tensor imaging; FDG-PET, <sup>18</sup>F-fluorodeoxyglucose positron emission tomography; H-MRSI, proton magnetic resonance spectroscopic imaging; MRI-VBM, magnetic resonance imaging–voxel-based morphometry; SAI, short-latency afferent inhibition; SPECT, single-photon emission computed tomography; TMS, transcranial magnetic stimulation.

callosum, paralleling prior reports of reduced callosal fractional anisotropy and cortical thickness of the corpus callosum<sup>56,64,66,67</sup>.

Functional imaging with <sup>18</sup>F-fluorodeoxyglucose positron emission tomography (FDG-PET) and SPECT has highlighted the involvement of frontal and temporal structures, even in the very initial phases of the disease <sup>28,68–72</sup>, possibly reflecting disease severity <sup>68</sup>.

Proton magnetic resonance spectroscopic imaging (H-MRSI) studies have shown a decreased N-acetyl aspartate/creatine ratio in the frontal and parietal cortices, centrum semiovale, and caudate nucleus, and there were significant correlations between clinical staging scale scores and H-MRSI abnormalities<sup>59,73,74</sup>.

As it clearly emerges, MRI is the modality of choice for identifying reported abnormalities in the clinical setting, such as frontal lobe and cerebellar atrophy<sup>4,75</sup>, white matter hyperintensities in parieto-occipital periventricular regions<sup>75</sup>, deep gray matter and hippocampal atrophy particularly in adult-onset patients<sup>54</sup>, and reduced midbrain-to-pons ratio<sup>59</sup>. However, specific neuroimaging findings are lacking in NPC, providing little aid in the clinical setting and diagnostic workup, highlighted by the absence of imaging tests in diagnostic criteria or from the NPC suspicion index<sup>76</sup>. Furthermore, these biomarkers have not been validated regarding accuracy in the differential diagnosis with other neurodegenerative diseases or compared with healthy controls; thus, specificity and sensitivity measures are currently unavailable. However, new imaging modalities have provided a solid basis for the development of biomarkers to understand disease pathophysiology and to monitor disease progression and response to treatments in the research setting.

#### Neurophysiology

As patients with NPC can experience any type of seizure (partial/focal, generalized, absence, myoclonic, or tonic-clonic), which vary in intensity and frequency, electroencephalography (EEG) should be used for confirmation and for differentiating epilepsy from cataplexy<sup>3,77–83</sup>. Patients who develop severe epilepsy generally have a worse prognosis and reduced life span compared with patients who are seizure-free<sup>3</sup>.

Neurophysiological evaluation with evoked potentials has shown the involvement of central tracts in most patients with NPC. Indeed, somatosensory evoked potentials have been shown to be impaired in patients with NPC, particularly in the lower limbs<sup>84,85</sup>, whereas brain auditory evoked potentials showed a bilateral absence of most waves, highlighting the involvement of auditory pathways ranging from the auditory nerve to the midbrain<sup>84–87</sup>.

Pyramidal involvement<sup>85,88</sup> and abnormalities of visual evoked potentials<sup>85</sup> have been reported in patients with infantile-onset NPC but appear only at advanced stages of disease<sup>84</sup>. Peripheral neuropathy has also been reported as a rare manifestation in patients with NPC<sup>3,68,89,90</sup>.

Recently, transcranial magnetic stimulation (TMS) pairedpulse paradigms have been used to evaluate cortical excitability and intracortical connectivity measures. In this context, an impairment in short-latency afferent inhibition (SAI), a measurement of sensorimotor integration, has been observed in NPC. SAI is thought to be mediated largely by central cholinergic transmission and has been shown to be impaired in patients with AD91,92, further supporting the link between NPC and AD60,93,94. Moreover, long-term potentiation (LTP)-like cortical plasticity, evaluated with a paired associative stimulation protocol, has been shown to be impaired in patients with NPC and in a symptomatic heterozygous carrier, confirming previous reports of impaired hippocampal synaptic plasticity in Npc1-mutant mice95. The impairment in LTP-like cortical plasticity has also been observed in patients with AD and FTD, further highlighting the possible parallelism between these disorders 96,97. Interestingly, after 12 months of treatment with miglustat, a considerable improvement in SAI and LTP-like plasticity was observed in patients with NPC<sup>60</sup>.

Only a limited decrease in short-interval intracortical inhibition, a marker of GABAAergic transmission, and intracortical facilitation, a marker of glutamatergic transmission, has been observed. Long-interval intracortical inhibition, reflecting GABABergic transmission, was reported to be within normal range<sup>60</sup>.

In conclusion, neurophysiological tests still fall short in providing invaluable information for the diagnostic workup of NPC diagnosis because findings are not specific for NPC and may not be found in all cases of NPC. EEG remains the exam of choice for confirming and differentiating epilepsy from cataplexy and for monitoring response to antiepileptic drug treatment.

However, in the research setting, a new series of non-invasive tests have shown a selective impairment of specific intracortical circuits that, if confirmed in a wider population of patients, might provide valuable information on disease pathophysiology and disease progression and possibly be used to monitor response to therapeutic interventions.

#### Neuropathology

Although cholesterol is able to flow freely through most cellular membranes, it cannot exit from lysosomes without the aid of NPC1 and NPC2 proteins. The so-called "hydrophobic handoff" model has been proposed<sup>98</sup>, and recent studies with structural biology approaches, such as crystallography and cryo–electron microscopy, have corroborated this model hypothesis, highlighting how the luminal NPC2 protein picks up cholesterol from endocytosed cholesterol as well as from the significant lipid content present in the lumen of degradative lysosomes, eventually interacting with the membrane-bound NPC1 protein lo-19. However, how cholesterol is transferred across the membrane by the NPC1 protein is still a matter of debate<sup>99</sup>.

Both NPC1 and NPC2 proteins are involved in the trafficking of low-density lipoprotein (LDL)-derived cholesterol from the lysosome to the cellular membranes of the endoplasmic reticulum, Golgi apparatus, and plasma membrane<sup>17,100</sup>. The decrease of cholesterol levels in the endoplasmic reticulum consequently enhances the synthesis and uptake of cholesterol by the

sterol response element-binding protein pathway, leading to the accumulation of cholesterol and other lipids in many types of cells, including lipid-laden macrophages (called foam cells) and neuronal and glial cells<sup>101-103</sup>.

Compared with most other lipid storage disorders, NPC does not arise from defective substrate degradation but from the impairment of LDL-derived cholesterol export out of the lysosome, followed by the disruption of lipid homeostasis <sup>103</sup>, affecting multiple cellular functions such as lysosomal calcium homeostasis <sup>104</sup>, oxidative stress <sup>105,106</sup>, Rab-mediated vesicle trafficking <sup>107,108</sup>, or fusion of lysosomes <sup>109</sup>, leading to impaired autophagy <sup>110,111</sup>. The accumulation of lipids in the central nervous system causes neuronal distension, axonal swelling, and the formation of axonal spheroids <sup>112–115</sup>.

NPC is also characterized by the accumulation of  $\beta$ -amyloid<sup>116,117</sup> and neurofibrillary tangles<sup>24,118–122</sup>, which are immunologically and ultrastructurally similar to those seen in AD<sup>123</sup>. Indeed, in one of the first descriptions of NPC neuropathology, a widespread neurofibrillary degeneration with a distribution similar to that of advanced AD was reported<sup>124</sup>. However, subsequent studies highlighted a somewhat different distribution of neurofibrillary tangles in NPC compared with AD<sup>71</sup>; there was a primary involvement of subcortical structures, including hippocampus, thalamus, and striatum in NPC<sup>24,118–120</sup>, and a more cortical distribution in AD<sup>125–127</sup>. Neurofibrillary tangles in NPC tend to be associated with lipid accumulations in swelling neurons, possibly suggesting a triggering effect of intracellular accumulations on tau aggregation<sup>71,118</sup>.

Beyond cholesterol pathway NPC and AD have other similarities, which involve AD pathogenesis  $^{128}$ . This parallelism between NPC and AD is further strengthened by the observation that cholesterol levels may modulate the processing of amyloid precursor protein  $^{129}$  and accumulation of  $\beta$ -amyloid  $^{130}$  is supported by the disease-modifying effect of the  $\epsilon 4$  isoform of apolipoprotein E on disease progression in both NPC and AD  $^{131-133}$ . Moreover, a possible effect of mutations in the NPC genes as AD risk factors has been speculated  $^{133,134}$ .

Just recently, a novel link with another intracellular protein-opathy has been established. Indeed, both in NPC mouse and in a human neuronal model of the disease, an altered expression or mislocalization of the TAR-DNA binding protein 43 (TDP-43) or both were reported  $^{135}$ . From a functional point of view, the TDP-43 mislocalization observed in human experimental neuronal models of NPC was associated with specific alterations in TDP-43 controlled genes. Most interestingly, N-acetyl-cysteine or 2-hydroxypropyl- $\beta$ -cyclodextrin may partially restore TDP-43 metabolism  $^{135}$ . TDP-43 inclusions have been reported as the main pathological signature of FTD due to *C9orf72* and *GRN* mutations  $^{136}$  and are also described in AD cases  $^{137}$ .

Whereas  $\beta$ -amyloid and  $\alpha$ -synuclein may accumulate biochemically in the NPC brain, senile plaques or Lewy bodies are not

characteristic of the disease, and further studies are needed to assess the possible overlap between these neurodegenerative disorders. As highlighted above, these speculations have not been validated and replicated in larger studies and thus a cautious interpretation is warranted.

#### **Treatments**

Miglustat, a small iminosugar molecule that reversibly inhibits glycosphingolipid synthesis, has been licensed in the European Union and elsewhere for the treatment of progressive neurological manifestations of NPC in both children and adults<sup>3,13,83,106</sup>. It has been shown to stabilize or improve certain neurological manifestations in six clinical trials<sup>28,88,138–142</sup>, none of which is randomized or placebo-controlled, and to partially restore neurophysiological markers of cholinergic impairment (such as SAI) and LTP-like cortical plasticity<sup>60,95</sup>.

The effect of cholesterol-lowering agents on hepatic and plasma cholesterol in NPC has been assessed with dimethyl sulfoxide, nicotinic acid, lovastatin, cholestyramine, and combinations of the above drugs. The treatment effects on total cholesterol varied depending on the drug combinations and overall improved with the number of drugs. However, efficacy for neurological outcomes was not reported, and safety findings discouraged widespread application of cholesterol-lowering agents to patients with NPC143,144. Many other therapies, including 2-hydroxypropyl-β-cyclodextrin (NCT02912793, NCT02939547, NCT01747135, and NCT02534844), arimoclomol (NCT02612129), vorinostat (NCT02124083), lithium carbonate (NCT03201627), and δ-tocopherol, are currently under clinical investigation for NPC145.

Although the mechanism of action of 2-hydroxypropyl-β-cyclodextrin is not fully understood, studies in animal models have shown reduced cholesterol and sphingolipid storage and liver function improvement, lower degree of neurodegeneration, and better survival following intravenous, subcutaneous, intracerebroventricular, or intrathecal administration, for both *NPC1* and *NPC2* mutations<sup>146–154</sup>.

Just recently, a phase 1-2 clinical trial with monthly 2-hydroxypropyl-β-cyclodextrin was performed on 14 patients with NPC and showed slowed disease progression, in particular in ambulation, cognition, and speech, with an acceptable safety profile<sup>155</sup>. Since 2-hydroxypropyl-β-cyclodextrin does not efficiently cross the blood-brain barrier<sup>156</sup> and high-dose systemic delivery can be associated with pulmonary toxicity149,157, lumbar intrathecal administration is the route of choice but has common adverse events, such as post-lumbar puncture headache, reported in 64% of cases 155. At doses above 600 mg, unexpected adverse events included post-administration unsteadiness and fatigue, which were transient and typically occurred 24 to 72 hours after dosing<sup>155</sup>. Ototoxicity, with mid- to high-frequency hearing loss, an expected adverse event, was documented in all participants and was probably due to outer hair cell loss<sup>155,158</sup>.

The main drawback of this approach is due to the route of administration; owing to the ability of molecular chaperones of the heat shock protein 70 (HSP70) family to protect pathologically challenged cells, HSP70-based therapies are emerging as attractive treatment options for many degenerative diseases 159-163, including lipid storage disorders due to their direct interaction with lysosomes 162,164, and for the proper folding and activity of the NPC1 protein 165,166. In this view, arimoclomol and small-molecule HSP70 co-inducer have been tested in a number of clinical trials 167,168 and are currently under investigation for the treatment of NPC.

Vorinostat, currently used for cutaneous T-cell lymphoma, is a histone deacetylase inhibitor that was able to increase NPC1 protein and decrease unesterified cholesterol deposits 169–173. Indeed, in selected genetic disorders, histone deacetylase inhibitors have been shown to induce histone modifications that not only can result in increased or decreased transcriptional expression of mutated genes 174 but also confer indirect benefits through acetylation of non-histone proteins, such as transcription factors and heat shock proteins, that modulate chaperones and proteostatic networks 169,174–176. Interestingly, however, treatment of *NPC2*-deficient human fibroblasts with a histone deacetylase inhibitor did not reduce cholesterol storage in lysosomes and late endosomes 171.

FTY720/fingolimod, an inhibitor of class I histone deacetylases used for the treatment of multiple sclerosis, has been shown to increase the expression of NPC1 and NPC2 in human *NPC1*-mutant fibroblasts and to significantly reduce the accumulation of cholesterol and glycosphingolipids<sup>177</sup>.

Other therapeutic approaches—including several that use human stem cells, such as hematopoietic stem cell transplantation (NCT00668564, NCT00730314, and NCT01372228), human placental-derived stem cell transplantation (NCT01586455), and intrathecal umbilical cord blood—derived oligodendrocyte-like cells (NCT02254863)—are currently under development for the treatment of NPC.

In regard to *NPC2* mutations, given that the NPC2 protein is soluble, secreted, and recaptured, there is a rationale supporting early hematopoietic stem cell transplantation<sup>178,179</sup>.

Other attractive approaches, evaluated in preclinical models of disease, include the systemic delivery of adeno-associated virus vectors to *NPC1*<sup>-/-</sup> mice to increase the expression of a therapeutic *NPC1* transgene, which has resulted in an improved clinical appearance, delayed weight loss,

significantly increased life span, reduced cholesterol storage, and decreased cerebellar Purkinje cell degeneration compared with untreated  $NPC1^{-l-}$  mice<sup>180,181</sup>.

#### **Conclusions**

NPC represents an autosomal recessive disorder with extensive biochemical, molecular, and clinical variability, which probably results in an underestimation of the burden of NPC cases worldwide. The relatively low incidence of the disease increases the difficulties in developing high-quality observational studies or randomized clinical trials.

As outlined above, recent studies have tried to shed light on the pathophysiology of this disease, further underlying its complex nature. Nevertheless, numerous biomarkers reflecting disease pathogenesis have emerged, thus representing a useful aid to diagnose disease or to evaluate disease progression and response to therapeutic interventions. Indeed, in this view, imaging and neurophysiological markers have been shown to reflect disease severity and to respond to disease-modifying treatments.

Intriguingly, a close parallelism has been observed between NPC and other neurodegenerative disorders, highlighting the possible involvement of multiple, but intertwined, disease pathways. Thus, unravelling the connection between neurofibrillary tangles, TDP-43 pathology, and neurodegeneration could result in important advances not only for NPC but also for AD and FTD/amyotrophic lateral sclerosis.

Diagnosing NPC represents a challenge for physicians, and delays in diagnosis and ensuing miglustat treatment and eventually future disease-specific interventions may affect disease outcomes because of irreversible anatomical damage and progressive neurodegeneration<sup>182</sup>. In this view, a prompt diagnosis is essential, and development of clinical tools, such as suspicion index to provide a risk prediction score<sup>76</sup>, along with instrumental diagnostic and prognostic markers, is mandatory. Efforts to increase awareness of NPC among clinicians are still needed, but the recent development of rapid and relatively simple instrumental and laboratory tests should improve the diagnostic and prognostic approach to NPC<sup>49</sup>.

#### Competing interests

The authors declare that they have no competing interests.

#### Grant information

The author(s) declare that no grants were involved in supporting this work.

#### References

- Carstea ED, Morris JA, Coleman KG, et al.: Niemann-Pick C1 disease gene: homology to mediators of cholesterol homeostasis. Science. 1997; 277(5323): 228–31
  - PubMed Abstract | Publisher Full Text
- Naureckiene S, Sleat DE, Lackland H, et al.: Identification of HE1 as the second gene of Niemann-Pick C disease. Science. 2000; 290(5500): 2298–301.
   PubMed Abstract | Publisher Full Text
- Patterson MC, Hendriksz CJ, Walterfang M, et al.: Recommendations for the diagnosis and management of Niemann-Pick disease type C: an update. Mol Genet Metab. 2012; 106(3): 330–44.
   PubMed Abstract
- Vanier MT: Niemann-Pick disease type C. Orphanet J Rare Dis. 2010; 5: 16. PubMed Abstract | Publisher Full Text | Free Full Text
- 5. Jahnova H, Dvorakova L, Vlaskova H, et al.: Observational, retrospective study

F1000 recommended

of a large cohort of patients with Niemann-Pick disease type C in the Czech Republic: a surprisingly stable diagnostic rate spanning almost 40 years.

Orphanet J Rare Dis. 2014; 9: 140.

PubMed Abstract | Publisher Full Text | Free Full Text

- 6. F Wassif CA, Cross JL, Iben J, et al.: High incidence of unrecognized visceral/ neurological late-onset Niemann-Pick disease, type C1, predicted by analysis of massively parallel sequencing data sets. Genet Med. 2016; 18(1): 41–8. PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- Pentchev PG, Comly ME, Kruth HS, et al.: Group C Niemann-Pick disease: faulty regulation of low-density lipoprotein uptake and cholesterol storage in cultured fibroblasts. FASEB J. 1987; 1(1): 40–5.
   PubMed Abstract | Publisher Full Text
- Liscum L, Ruggiero RM, Faust JR: The intracellular transport of low density lipoprotein-derived cholesterol is defective in Niemann-Pick type C fibroblasts. J Cell Biol. 1989; 108(5): 1625–36.
   PubMed Abstract | Publisher Full Text | Free Full Text
- Pentchev PG, Comly ME, Kruth HS, et al.: A defect in cholesterol esterification in Niemann-Pick disease (type C) patients. Proc Natl Acad Sci U S A. 1985; 82(23): 8247–51.

PubMed Abstract | Free Full Text

- Li X, Lu F, Trinh MN, et al.: 3.3 Å structure of Niemann-Pick C1 protein reveals insights into the function of the C-terminal luminal domain in cholesterol transport. Proc Natl Acad Sci U S A. 2017; 114(34): 9116–21.
   PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- Poongavanam V, Kongsted J, Wüstner D: Computational Analysis of Sterol Ligand Specificity of the Niemann Pick C2 Protein. Biochemistry. 2016; 55(36): 5165–79.
  - PubMed Abstract | Publisher Full Text | F1000 Recommendation
- Li X, Saha P, Li J, et al.: Clues to the mechanism of cholesterol transfer from the structure of NPC1 middle lumenal domain bound to NPC2. Proc Natl Acad Sci U S A. 2016; 113(36): 10079–84.
   PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- Li X, Wang J, Coutavas E, et al.: Structure of human Niemann-Pick C1 protein. Proc Natl Acad Sci U S A. 2016; 113(29): 8212–7.
   PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- Gong X, Qian H, Zhou X, et al.: Structural Insights into the Niemann-Pick C1 (NPC1)-Mediated Cholesterol Transfer and Ebola Infection. Cell. 2016; 165(6): 1467–78.

PubMed Abstract | Publisher Full Text

 Zhao Y, Ren J, Harlos K, et al.: Structure of glycosylated NPC1 luminal domain C reveals insights into NPC2 and Ebola virus interactions. FEBS Lett. 2016; 590(5): 605–12.

PubMed Abstract | Publisher Full Text | Free Full Text

- Wang ML, Motamed M, Infante RE, et al.: Identification of surface residues on Niemann-Pick C2 essential for hydrophobic handoff of cholesterol to NPC1 in Iyosomes. Cell Metab. 2010; 12(2): 166–73.
   PubMed Abstract | Publisher Full Text | Free Full Text
- Kwon HJ, Abi-Mosleh L, Wang ML, et al.: Structure of N-terminal domain of NPC1 reveals distinct subdomains for binding and transfer of cholesterol. Cell. 2009; 137(7): 1213–24.
   PublMed Abstract | Publisher Full Text | Free Full Text
- Xu S, Benoff B, Liou HL, et al.: Structural basis of sterol binding by NPC2, a lysosomal protein deficient in Niemann-Pick type C2 disease. J Biol Chem. 2007; 282(32): 23525–31.

PubMed Abstract | Publisher Full Text | Free Full Text

- Friedland N, Liou HL, Lobel P, et al.: Structure of a cholesterol-binding protein deficient in Niemann-Pick type C2 disease. Proc Natl Acad Sci U S A. 2003; 100(5): 2512–7.
  - PubMed Abstract | Publisher Full Text | Free Full Text
- Sévin M, Lesca G, Baumann N, et al.: The adult form of Niemann-Pick disease type C. Brain. 2007; 130(Pt 1): 120–33.
   PubMed Abstract | Publisher Full Text | F1000 Recommendation
- Millat G, Chikh K, Naureckiene S, et al.: Niemann-Pick disease type C: spectrum
  of HE1 mutations and genotype/phenotype correlations in the NPC2 group. Am
  J Hum Genet. 2001; 69(5): 1013–21.
   PubMed Abstract | Publisher Full Text | Free Full Text
- Park WD, O'Brien JF, Lundquist PA, et al.: Identification of 58 novel mutations in Niemann-Pick disease type C: correlation with biochemical phenotype and importance of PTC1-like domains in NPC1. Hum Mutat. 2003; 22(4): 313–25.
   PubMed Abstract | Publisher Full Text
- Griese M, Brasch F, Aldana VR, et al.: Respiratory disease in Niemann-Pick type C2 is caused by pulmonary alveolar proteinosis. Clin Genet. 2010; 77(2): 119–30.
   PubMed Abstract | Publisher Full Text
- Klünemann HH, Elleder M, Kaminski WE, et al.: Frontal lobe atrophy due to a mutation in the cholesterol binding protein HE1/NPC2. Ann Neurol. 2002; 52(6): 743-9.
   PubMed Abstract | Publisher Full Text
- 25. Chikh K, Rodriguez C, Vey S, et al.: Niemann-Pick type C disease: subcellular location and functional characterization of NPC2 proteins with naturally

- occurring missense mutations. *Hum Mutat.* 2005; **26**(1): 20–8. PubMed Abstract | Publisher Full Text
- Yanjanin NM, Vélez JI, Gropman A, et al.: Linear clinical progression, independent of age of onset, in Niemann-Pick disease, type C. Am J Med Genet B Neuropsychiatr Genet. 2010; 153B(1): 132–40.
   PubMed Abstract | Publisher Full Text | Free Full Text
- Pineda M, Wraith JE, Mengel E, et al.: Miglustat in patients with Niemann-Pick disease Type C (NP-C): a multicenter observational retrospective cohort study. Mol Genet Metab. 2009; 98(3): 243-9.
   PubMed Abstract | Publisher Full Text
- Pineda M, Perez-Poyato MS, O'Callaghan M, et al.: Clinical experience with miglustat therapy in pediatric patients with Niemann-Pick disease type C: a case series. Mol Genet Metab. 2010; 99(4): 358–66.
   PubMed Abstract | Publisher Full Text
- Iturriaga C, Pineda M, Fernández-Valero EM, et al.: Niemann-Pick C disease in Spain: clinical spectrum and development of a disability scale. J Neurol Sci. 2006; 249(1): 1–6.
   PubMed Abstract | Publisher Full Text
- Stampfer M, Theiss S, Amraoui Y, et al.: Niemann-Pick disease type C clinical database: cognitive and coordination deficits are early disease indicators. Orphanet J Rare Dis. 2013; 8: 35.
   PubMed Abstract | Publisher Full Text | Free Full Text
- Jiang X, Sidhu R, Porter FD, et al.: A sensitive and specific LC-MS/MS method for rapid diagnosis of Niemann-Pick C1 disease from human plasma. J Lipid Res. 2011; 52(7): 1435–45.
   PubMed Abstract | Publisher Full Text | Free Full Text
- Porter FD, Scherrer DE, Lanier MH, et al.: Cholesterol oxidation products are sensitive and specific blood-based biomarkers for Niemann-Pick C1 disease. Sci Transl Med. 2010; 2(56): 56ra81.
   PubMed Abstract | Publisher Full Text | Free Full Text
- Vanier MT, Rodriguez-Lafrasse C, Rousson R, et al.: Type C Niemann-Pick disease: spectrum of phenotypic variation in disruption of intracellular LDLderived cholesterol processing. Biochim Biophys Acta. 1991; 1096(4): 328–37.
   PubMed Abstract | Publisher Full Text
- Vanier MT, Suzuki K: Recent advances in elucidating Niemann-Pick C disease. Brain Pathol. 1998; 8(1): 163–74.
   PubMed Abstract | Publisher Full Text
- Argoff CE, Comly ME, Blanchette-Mackie J, et al.: Type C Niemann-Pick disease: cellular uncoupling of cholesterol homeostasis is linked to the severity of disruption in the intracellular transport of exogenously derived cholesterol. Biochim Biophys Acta. 1991; 1096(4): 319–27.
   PubMed Abstract | Publisher Full Text
- Klinke G, Rohrbach M, Giugliani R, et al.: LC-MS/MS based assay and reference intervals in children and adolescents for oxysterols elevated in Niemann-Pick diseases. Clin Biochem. 2015; 48(9): 596–602.
   PubMed Abstract | Publisher Full Text
- Pajares S, Arias A, García-Villoria J, et al.: Cholestane-3β,5α,6β-triol: high levels in Niemann-Pick type C, cerebrotendinous xanthomatosis, and lysosomal acid lipase deficiency. J Lipid Res. 2015; 56(10): 1926–35.
   PubMed Abstract | Publisher Full Text | Free Full Text
- Reunert J, Fobker M, Kannenberg F, et al.: Rapid Diagnosis of 83 Patients with Niemann Pick Type C Disease and Related Cholesterol Transport Disorders by Cholestantriol Screening. EBioMedicine. 2015; 4: 170–5.
   PubMed Abstract | Publisher Full Text | Free Full Text
- Polo G, Burlina A, Furlan F, et al.: High level of oxysterols in neonatal cholestasis: a pitfall in analysis of biochemical markers for Niemann-Pick type C disease. Clin Chem Lab Med. 2016; 54(7): 1221–9.
   PubMed Abstract | Publisher FullText
- NP-C Guidelines Working Group, Wraith JE, Baumgartner MR, et al.: Recommendations on the diagnosis and management of Niemann-Pick disease type C. Mol Genet Metab. 2009; 98(1–2): 152–165.
   PubMed Abstract | Publisher Full Text
- Sun X, Marks DL, Park WD, et al.: Niemann-Pick C variant detection by altered sphingolipid trafficking and correlation with mutations within a specific domain of NPC1. Am J Hum Genet. 2001; 68(6): 1361–72.
   PubMed Abstract | Publisher Full Text | Free Full Text
- Fernandez-Valero EM, Ballart A, Iturriaga C, et al.: Identification of 25 new mutations in 40 unrelated Spanish Niemann-Pick type C patients: genotypephenotype correlations. Clin Genet. 2005; 68(3): 245–54.
   PubMed Abstract | Publisher Full Text
- Macías-Vidal J, Rodríguez-Pascau L, Sánchez-Ollé G, et al.: Molecular analysis
  of 30 Niemann-Pick type C patients from Spain. Clin Genet. 2011; 80(1): 39–49.
  PubMed Abstract | Publisher Full Text
- Kruth HS, Comly ME, Butler JD, et al.: Type C Niemann-Pick disease. Abnormal metabolism of low density lipoprotein in homozygous and heterozygous fibroblasts. J Biol Chem. 1986; 261(35): 16769–16774.
   PubMed Abstract
- 45. Vanier MT, Latour P: Laboratory diagnosis of Niemann-Pick disease type C: the filipin staining test. Methods Cell Biol. 2015; 126: 357–75.

  PubMed Abstract | Publisher Full Text | F1000 Recommendation
- Wortmann SB, Vaz FM, Gardeitchik T, et al.: Mutations in the phospholipid remodeling gene SERAC1 impair mitochondrial function and intracellular

cholesterol trafficking and cause dystonia and deafness. *Nat Genet.* 2012; **44**(7): 797–802.

PubMed Abstract | Publisher Full Text

47. Platt FM, Wassif C, Colaco A, et al.: Disorders of cholesterol metabolism and their unanticipated convergent mechanisms of disease. Annu Rev Genomics Hum Genet. 2014; 15: 173–94.

PubMed Abstract | Publisher Full Text

 Sechi A, Dardis A, Zampieri S, et al.: Effects of miglustat treatment in a patient affected by an atypical form of Tangier disease. Orphanet J Rare Dis. 2014; 9: 143

PubMed Abstract | Publisher Full Text | Free Full Text

- 49. Vanier MT, Gissen P, Bauer P, et al.: Diagnostic tests for Niemann-Pick disease type C (NP-C): A critical review. Mol Genet Metab. 2016; 118(4): 244–54. PubMed Abstract | Publisher Full Text | F1000 Recommendation
- 50. Jiang X, Sidhu R, Mydock-McGrane L, et al.: Development of a bile acid-based newborn screen for Niemann-Pick disease type C. Sci Transl Med. 2016; 8(337): 337ra63.
  PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- Platt FM, Neises GR, Dwek RA, et al.: N-butyldeoxynojirimycin is a novel inhibitor of glycolipid biosynthesis. J Biol Chem. 1994; 269(11): 8362–5.
   PubMed Abstract
- Patterson MC, Clayton P, Gissen P, et al.: Recommendations for the detection and diagnosis of Niemann-Pick disease type C: An update. Neurol Clin Pract. 2017; 7(6): 499–511.

PubMed Abstract | Publisher Full Text | Free Full Text

- 53. F Bowman EA, Walterfang M, Abel L, et al.: Longitudinal changes in cerebellar and subcortical volumes in adult-onset Niemann-Pick disease type C patients treated with miglustat. J Neurol. 2015; 262(9): 2106–14.
  PubMed Abstract | Publisher Full Text | F1000 Recommendation
- Walterfang M, Patenaude B, Abel LA, et al.: Subcortical volumetric reductions in adult Niemann-Pick disease type C: a cross-sectional study. AJNR Am J Neuroradiol. 2013; 34(7): 1334–40.
   PubMed Abstract | Publisher Full Text
- Zaaraoui W, Crespy L, Rico A, et al.: In vivo quantification of brain injury in adult Niemann-Pick Disease Type C. Mol Genet Metab. 2011; 103(2): 138–41.
   PubMed Abstract
- Walterfang M, Fahey M, Desmond P, et al.: White and gray matter alterations in adults with Niemann-Pick disease type C: a cross-sectional study. Neurology. 2010; 75(1): 49–56.

PubMed Abstract | Publisher Full Text

 Terbeek J, Latour P, Van Laere K, et al.: Abnormal dopamine transporter imaging in adult-onset Niemann-Pick disease type C. Parkinsonism Relat Disord. 2017; 36: 107–8.

PubMed Abstract | Publisher Full Text

- Walterfang M, Abel LA, Desmond P, et al.: Cerebellar volume correlates with saccadic gain and ataxia in adult Niemann-Pick type C. Mol Genet Metab. 2013; 108(1): 85–9.
   PubMed Abstract
- Walterfang M, Macfarlane MD, Looi JC, et al.: Pontine-to-midbrain ratio indexes ocular-motor function and illness stage in adult Niemann-Pick disease type C. Eur J Neurol. 2012; 19(3): 462–7.
   PubMed Abstract | Publisher Full Text
- Benussi A, Cotelli MS, Cosseddu M, et al.: Preliminary Results on Long-Term Potentiation-Like Cortical Plasticity and Cholinergic Dysfunction After Miglustat Treatment in Niemann-Pick Disease Type C. JIMD Rep. 2017; 36: 19-27

PubMed Abstract | Publisher Full Text | Free Full Text

- Scheel M, Abegg M, Lanyon LJ, et al.: Eye movement and diffusion tensor imaging analysis of treatment effects in a Niemann-Pick Type C patient. Mol Genet Metab. 2010; 99(3): 291–5.
   PubMed Abstract | Publisher Full Text
- Trouard TP, Heidenreich RA, Seeger JF, et al.: Diffusion tensor imaging in Niemann-Pick Type C disease. Pediatr Neurol. 2005; 33(5): 325–30.
   PubMed Abstract | Publisher Full Text
- 63. F Masingue M, Adanyeguh I, Nadjar Y, et al.: Evolution of structural neuroimaging biomarkers in a series of adult patients with Niemann-Pick type C under treatment. Orphanet J Rare Dis. 2017; 12(1): 22. PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- Walterfang M, Fahey M, Abel L, et al.: Size and shape of the corpus callosum in adult Niemann-Pick type C reflects state and trait illness variables. AJNR Am J Neuroradiol. 2011; 32(7): 1340–6.

PubMed Abstract | Publisher Full Text

- Prasloski T, Rauscher A, MacKay AL, et al.: Rapid whole cerebrum myelin water imaging using a 3D GRASE sequence. Neuroimage. 2012; 63(1): 533–9.
   PubMed Abstract | Publisher Full Text
- 66. Lee R, Apkarian K, Jung ES, et al.: Corpus callosum diffusion tensor imaging and volume measures are associated with disease severity in pediatric Niemann-Pick disease type C1. Pediatr Neurol. 2014; 51(5): 669–674.e5. PubMed Abstract | Publisher Full Text | Free Full Text

- Palmeri S, Battisti C, Federico A, et al.: Hypoplasia of the corpus callosum in Niemann-Pick type C disease. Neuroradiology. 1994; 36(1): 20–2.
   PubMed Abstract | Publisher Full Text
- Benussi A, Alberici A, Premi E, et al.: Phenotypic heterogeneity of Niemann-Pick disease type C in monozygotic twins. J Neurol. 2015; 262(3): 642–7.
   PubMed Abstract | Publisher Full Text
- Battisti C, Tarugi P, Dotti MT, et al.: Adult onset Niemann-Pick type C disease: A clinical, neuroimaging and molecular genetic study. Mov Disord. 2003; 18(11): 1405–9.

PubMed Abstract | Publisher Full Text

 Walterlang M, Fietz M, Fahey M, et al.: The neuropsychiatry of Niemann-Pick type C disease in adulthood. J Neuropsychiatry Clin Neurosci. 2006; 18(2): 158–70.

PubMed Abstract | Publisher Full Text

- 71. Bergeron D, Poulin S, Laforce R Jr: Cognition and anatomy of adult Niemann-Pick disease type C: Insights for the Alzheimer field. Cogn Neuropsychol. 2017; 1–14.

  PubMed Abstract | Publisher Full Text | F1000 Recommendation
- Kumar A, Chugani HT: Niemann-Pick disease type C: unique 2-deoxy-2[18F] fluoro-D-glucose PET abnormality. Pediatr Neurol. 2011; 44(1): 57–60.
   PubMed Abstract | Publisher Full Text
- Tedeschi G, Bonavita S, Barton NW, et al.: Proton magnetic resonance spectroscopic imaging in the clinical evaluation of patients with Niemann-Pick type C disease. J Neurol Neurosurg Psychiatry. 1998; 65(1): 72–9.
   PubMed Abstract | Publisher Full Text | Free Full Text
- Galanaud D, Tourbah A, Lehéricy S, et al.: 24 month-treatment with miglustat
  of three patients with Niemann-Pick disease type C: follow up using brain
  spectroscopy. Mol Genet Metab. 2009; 96(2): 55–8.
   PubMed Abstract | Publisher Full Text
- Huang JY, Peng SF, Yang CC, et al.: Neuroimaging findings in a brain with Niemann-Pick type C disease. J Formos Med Assoc. 2011; 110(8): 537–42.
   PubMed Abstract | Publisher Full Text
- Wijburg FA, Sedel F, Pineda M, et al.: Development of a suspicion index to aid diagnosis of Niemann-Pick disease type C. Neurology. 2012; 78(20): 1560–7.
   PubMed Abstract | Publisher Full Text
- Vanier MT, Millat G: Niemann-Pick disease type C. Clin Genet. 2003; 64(4): 269–81.
   PubMed Abstract | Publisher Full Text

Zarowski M, Steinborn B, Gurda B, et al.: Treatment of cataplexy in Niemann-Pick disease type C with the use of miglustat. Eur J Paediatr Neurol. 2011; 15(1):

84–7.
PubMed Abstract | Publisher Full Text

- Smit LS, Lammers GJ, Catsman-Berrevoets CE: Cataplexy leading to the diagnosis of Niemann-Pick disease type C. Pediatr Neurol. 2006; 35(1): 82–4.
   PubMed Abstract | Publisher Full Text
- Philippart M, Engel J Jr, Zimmerman EG: Gelastic cataplexy in Niemann-Pick disease group C and related variants without generalized sphingomyelinase deficiency. Ann Neurol. 1983; 14(4): 492–3.
   PubMed Abstract | Publisher Full Text
- Koens LH, Kuiper A, Coenen MA, et al.: Ataxia, dystonia and myoclonus in adult patients with Niemann-Pick type C. Orphanet J Rare Dis. 2016; 11(1): 121.
   PubMed Abstract | Publisher Full Text | Free Full Text
- Canafoglia L, Bugiani M, Uziel G, et al.: Rhythmic cortical myoclonus in Niemann-Pick disease type C. Mov Disord. 2006; 21(9): 1453–6.
   PubMed Abstract | Publisher Full Text
- Piña-Aguilar RE, Vera-Loaiza A, Chacón-Camacho OF, et al.: Clinical and genetic characteristics of mexican patients with juvenile presentation of niemann-pick type C disease. Case Rep Neurol Med. 2014; 2014: 785890.
   PubMed Abstract | Publisher Full Text | Free Full Text
- 84. Flodice R, Dubbioso R, Topa A, et al.: Electrophysiological characterization of adult-onset Niemann-Pick type C disease. J Neurol Sci. 2015; 348(1–2): 262–5. PubMed Abstract | Publisher Full Text | F1000 Recommendation
- Higgins JJ, Patterson MC, Dambrosia JM, et al.: A clinical staging classification for type C Niemann-Pick disease. Neurology. 1992; 42(12): 2286–90.
   PubMed Abstract | Publisher Full Text
- King KA, Gordon-Salant S, Yanjanin N, et al.: Auditory phenotype of Niemann-Pick disease, type C1. Ear Hear. 2014; 35(1): 110–7.
   PubMed Abstract | Publisher Full Text | Free Full Text
- Pikus A: Audiologic Profile in Niemann-Pick C. Ann NY Acad Sci. 1991; 630: 313–4.

PubMed Abstract | Publisher Full Text

 Patterson MC, Vecchio D, Prady H, et al.: Miglustat for treatment of Niemann-Pick C disease: A randomised controlled study. Lancet Neurol. 2007; 6(9): 765–72

PubMed Abstract | Publisher Full Text

- Hahn AF, Gilbert JJ, Kwarciak C, et al.: Nerve biopsy findings in Niemann-Pick type II (NPC). Acta Neuropathol. 1994; 87(2): 149–54.
   PubMed Abstract | Publisher Full Text
- 90. Zafeiriou DI, Triantafyllou P, Gombakis NP, et al.: Niemann-pick type C disease

- associated with peripheral neuropathy. Pediatr Neurol. 2003; 29(3): 242–4. PubMed Abstract | Publisher Full Text
- Di Lazzaro V, Oliviero A, Tonali PA, et al.: Noninvasive in vivo assessment of cholinergic cortical circuits in AD using transcranial magnetic stimulation. Neurology. 2002; 59(3): 392–7.
   PubiMed Abstract | Publisher Full Text
- Benussi A, Padovani A, Borroni B: Transcranial Magnetic Stimulation in Alzheimer's Disease and Cortical Dementias. J Alzheimers Dis Parkinsonism. 2015; 05(03).
   Publisher Full Text
- Manganelli F, Dubbioso R, Iodice R, et al.: Central cholinergic dysfunction in the adult form of Niemann Pick disease type C: a further link with Alzheimer's disease? J Neurol. 2014; 261(4): 804–8.
   PubMed Abstract | Publisher Full Text
- Benussi A, Di Lorenzo F, Dell'Era V, et al.: Transcranial magnetic stimulation distinguishes Alzheimer disease from frontotemporal dementia. Neurology. 2017; 89(7): 665–72.
   PubMed Abstract | Publisher Full Text
- 95. D'Arcangelo G, Grossi D, Racaniello M, et al.: Miglustat Reverts the Impairment of Synaptic Plasticity in a Mouse Model of NPC Disease. Neural Plast. 2016; 2016: 3830424.

  PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- 96. Di Lorenzo F, Ponzo V, Bonni S, et al.: Long-term potentiation-like cortical plasticity is disrupted in Alzheimer's disease patients independently from age of onset. Ann Neurol. 2016; 80(2): 202–10.
- PubMed Abstract | Publisher Full Text | F1000 Recommendation
- Benussi A, Cosseddu M, Filareto I, et al.: Impaired long-term potentiation-like cortical plasticity in presymptomatic genetic frontotemporal dementia. Ann Neurol. 2016; 80(3): 472–6.
   PubMed Abstract | Publisher Full Text
- Brown MS, Goldstein JL: A receptor-mediated pathway for cholesterol homeostasis. Science. 1986; 232(4746): 34–47.
   PubMed Abstract | Publisher Full Text
- Pfeffer SR: Clues to NPC1-mediated cholesterol export from lysosomes. Proc Natl Acad Sci U S A. 2016; 113(29): 7941–3.
   PubMed Abstract | Publisher Full Text | Free Full Text
- Infante RE, Wang ML, Radhakrishnan A, et al.: NPC2 facilitates bidirectional transfer of cholesterol between NPC1 and lipid bilayers, a step in cholesterol egress from lysosomes. Proc Natl Acad Sci U S A. 2008; 105(40): 15287–92.
   PubMed Abstract | Publisher Full Text | Free Full Text
- Liscum L, Faust JR: Low density lipoprotein (LDL)-mediated suppression of cholesterol synthesis and LDL uptake is defective in Niemann-Pick type C fibroblasts. J Biol Chem. 1987; 262(35): 17002–8.
   PubMed Abstract
- Brown MS, Goldstein JL: The SREBP Pathway: regulation of cholesterol metabolism by proteolysis of a membrane-bound transcription factor. Cell. 1997; 89(3): 331-40.
   PubMed Abstract | Publisher Full Text
- 103. E Kuech EM, Brogden G, Naim HY: Alterations in membrane trafficking and pathophysiological implications in lysosomal storage disorders. Biochimie. 2016; 130: 152–62.
  - PubMed Abstract | Publisher Full Text | F1000 Recommendation
- Lloyd-Evans E, Morgan AJ, He X, et al.: Niemann-Pick disease type C1 is a sphingosine storage disease that causes deregulation of lysosomal calcium. Nat Med. 2008; 14(11): 1247–55.
   PubMed Abstract | Publisher Full Text
- 105. Fu R, Yanjanin NM, Bianconi S, et al.: Oxidative stress in Niemann-Pick disease, type C. Mol Genet Metab. 2010; 101(2–3): 214–8.
  PubMed Abstract | Publisher Full Text | Free Full Text
- Vázquez MC, Balboa E, Alvarez AR, et al.: Oxidative stress: a pathogenic mechanism for Niemann-Pick type C disease. Oxid Med Cell Longev. 2012; 2012: 205713.
  - PubMed Abstract | Publisher Full Text | Free Full Text
- Choudhury A, Dominguez M, Puri V, et al.: Rab proteins mediate Golgi transport
  of caveola-internalized glycosphingolipids and correct lipid trafficking in
  Niemann-Pick C cells. J Clin Invest. 2002; 109(12): 1541–50.
   PubMed Abstract | Publisher Full Text | Free Full Text
- Choudhury A, Sharma DK, Marks DL, et al.: Elevated endosomal cholesterol levels in Niemann-Pick cells inhibit rab4 and perturb membrane recycling. Mol Biol Cell. 2004; 15(10): 4500-11.
   PubMed Abstract | Publisher Full Text | Free Full Text
- Goldman SD, Krise JP: Niemann-Pick C1 functions independently of Niemann-Pick C2 in the initial stage of retrograde transport of membrane-impermeable lysosomal cargo. J Biol Chem. 2010; 285(7): 4983–94.
   PubMed Abstract | Publisher Full Text | Free Full Text
- Sarkar S, Carroll B, Buganim Y, et al.: Impaired autophagy in the lipid-storage disorder Niemann-Pick type C1 disease. Cell Rep. 2013; 5(5): 1302–15.
   PubMed Abstract | Publisher Full Text | Free Full Text
- Pacheco CD, Kunkel R, Lieberman AP: Autophagy in Niemann-Pick C disease is dependent upon Beclin-1 and responsive to lipid trafficking defects. Hum Mol Genet. 2007; 16(12): 1495–503.
   PubMed Abstract | Publisher Full Text

- March PA, Thrall MA, Brown DE, et al.: GABAergic neuroaxonal dystrophy and other cytopathological alterations in feline Niemann-Pick disease type C. Acta Neuropathol. 1997; 94(2): 164–72.
   PubMed Abstract | Publisher Full Text
- Zervas M, Dobrenis K, Walkley SU: Neurons in Niemann-Pick disease type C accumulate gangliosides as well as unesterified cholesterol and undergo dendritic and axonal alterations. J Neuropathol Exp Neurol. 2001; 60(1): 49–64.
   PubMed Abstract | Publisher Full Text
- Pressey SN, Smith DA, Wong AM, et al.: Early glial activation, synaptic changes and axonal pathology in the thalamocortical system of Niemann-Pick type C1 mice. Neurobiol Dis. 2012; 45(3): 1086–100.
   PubMed Abstract | Publisher Full Text | Free Full Text
- 115. Walkley SU, Baker HJ, Rattazzi MC, et al.: Neuroaxonal dystrophy in neuronal storage disorders: evidence for major GABAergic neuron involvement. J Neurol Sci. 1991; 104(1): 1–8. PubMed Abstract | Publisher Full Text
- Nixon RA: Niemann-Pick Type C Disease and Alzheimer's disease: the APPendosome connection fattens up. Am J Pathol. 2004; 164(3): 757–61.
   PubMed Abstract | Publisher Full Text | Free Full Text
- 117. Yamazaki T, Chang TY, Haass C, et al.: Accumulation and aggregation of amyloid beta-protein in late endosomes of Niemann-pick type C cells. J Biol Chem. 2001; 276(6): 4454–60.
  PubMed Abstract | Publisher Full Text
- Love S, Bridges LR, Case CP: Neurofibrillary tangles in Niemann—Pick disease type C. Brain. 1995; 118(Pt 1): 119–29.
   PubMed Abstract | Publisher Full Text
- Suzuki K, Parker CC, Pentchev PG, et al.: Neurofibrillary tangles in Niemann-Pick disease type C. Acta Neuropathol. 1995; 89(3): 227–38.
   PubMed Abstract | Publisher Full Text
- Chiba Y, Komori H, Takei S, et al.: Niemann-Pick disease type C1 predominantly involving the frontotemporal region, with cortical and brainstem Lewy bodies: an autopsy case. Neuropathology. 2014; 34(1): 49–57.
   PubMed Abstract | Publisher Full Text
- Ohm TG, Treiber-Held S, Distl R, et al.: Cholesterol and tau protein--findings in Alzheimer's and Niemann Pick C's disease. Pharmacopsychiatry. 2003; 36(Suppl 2): S120–6.
   PubMed Abstract | Publisher Full Text
- Distl R, Treiber-Held S, Albert F, et al.: Cholesterol storage and tau pathology in Niemann-Pick type C disease in the brain. J Pathol. 2003; 200(1): 104–11.
   PubMed Abstract | Publisher Full Text
- 123. Auer IA, Schmidt ML, Lee VM, et al.: Paired helical filament tau (PHFtau) in Niemann-Pick type C disease is similar to PHFtau in Alzheimer's disease. Acta Neuropathol. 1995; 90(6): 547-51. PubMed Abstract | Publisher Full Text
- 124. Horoupian DS, Yang SS: Paired helical filaments in neurovisceral lipidosis (juvenile dystonic lipidosis). Ann Neurol. 1978; 4(5): 404–11.
  PubMed Abstract | Publisher Full Text
- 125. F Braak H, Alafuzoff I, Arzberger T, et al.: Staging of Alzheimer disease-associated neurofibrillary pathology using paraffin sections and immunocytochemistry. Acta Neuropathol. 2006; 112(4): 389–404.
  PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- 126. Brier MR, Gordon B, Friedrichsen K, et al.: Tau and A

  ß imaging, CSF measures, and cognition in Alzheimer's disease. Sci Transl Med. 2016; 8(338): 338ra66.
  PubMed Abstract | Publisher Full Text | Free Full Text
- 127. F Schöll M, Lockhart SN, Schonhaut DR, et al.: PET Imaging of Tau Deposition in the Aging Human Brain. Neuron. 2016; 89(5): 971–82.
  PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- Liu CC, Liu CC, Kanekiyo T, et al.: Apolipoprotein E and Alzheimer disease: risk, mechanisms and therapy. Nat Rev Neurol. 2013; 9(2): 106–18.
   PubMed Abstract | Publisher Full Text | Free Full Text
- Bodovitz S, Klein WL: Cholesterol modulates alpha-secretase cleavage of amyloid precursor protein. J Biol Chem. 1996; 271(8): 4436–40.
   PubMed Abstract | Publisher Full Text
- Simons M, Keller P, De Strooper B, et al.: Cholesterol depletion inhibits the generation of beta-amyloid in hippocampal neurons. Proc Natl Acad Sci U S A. 1998; 95(11): 6460-4.
   PubMed Abstract | Free Full Text
- I31. Fu R, Yanjanin NM, Elrick MJ, et al.: Apolipoprotein E genotype and neurological disease onset in Niemann-Pick disease, type C1. Am J Med Genet A. 2012; 158A(11): 2775–80.
  PubMed Abstract | Publisher Full Text | Free Full Text
- Corder EH, Saunders AM, Strittmatter WJ, et al.: Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. Science. 1993; 261(5123): 921–3.
   PubMed Abstract | Publisher Full Text
- 133. Malnar M, Hecimovic S, Mattsson N, et al.: Bidirectional links between Alzheimer's disease and Niemann-Pick type C disease. Neurobiol Dis. 2014; 72(Pt A): 37–47. PubMed Abstract | Publisher Full Text
- Kresojević N, Dobričić V, Svetel M, et al.: Mutations in Niemann Pick type C gene are risk factor for Alzheimer's disease. Med Hypotheses. 2014; 83(5): 559–62.
   PubMed Abstract | Publisher Full Text

- 135. F Dardis A, Zampieri S, Canterini S, et al.: Altered localization and functionality of TAR DNA Binding Protein 43 (TDP-43) in niemann-pick disease type C. Acta Neuropathol Commun. 2016; 4(1): 52.

  PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- 136. Benussi A, Padovani A, Borroni B: Phenotypic Heterogeneity of Monogenic Frontotemporal Dementia. Front Aging Neurosci. 2015; 7: 171. PubMed Abstract | Publisher Full Text | Free Full Text
- Josephs KA, Murray ME, Whitwell JL, et al.: Updated TDP-43 in Alzheimer's disease staging scheme. Acta Neuropathol. 2016; 131(4): 571–85.
   PubMed Abstract | Publisher Full Text
- Butters TD, Dwek RA, Platt FM: Inhibition of glycosphingolipid biosynthesis: application to lysosomal storage disorders. Chem Rev. 2000; 100(12): 4683–96.
   PubMed Abstract | Publisher Full Text
- Patterson MC, Vecchio D, Jacklin E, et al.: Long-term miglustat therapy in children with Niemann-Pick disease type C. J Child Neurol. 2010; 25(3): 300–5.
   PubMed Abstract | Publisher Full Text
- 140. Wraith JE, Vecchio D, Jacklin E, et al.: Miglustat in adult and juvenile patients with Niemann-Pick disease type C: long-term data from a clinical trial. Mol Genet Metab. 2010; 99(4): 351–7. PubMed Abstract | Publisher Full Text
- Héron B, Valayannopoulos V, Baruteau J, et al.: Miglustat therapy in the French cohort of paediatric patients with Niemann-Pick disease type C. Orphanet J Rare Dis. 2012; 7: 36.
   PublMed Abstract | Publisher Full Text | Free Full Text
- 142. F Fecarotta S, Romano A, Della Casa R, et al.: Long term follow-up to evaluate the efficacy of miglustat treatment in Italian patients with Niemann-Pick disease type C. Orphanet J Rare Dis. 2015; 10: 22. PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- Patterson MC, Di Bisceglie AM, Higgins JJ, et al.: The effect of cholesterollowering agents on hepatic and plasma cholesterol in Niemann-Pick disease type C. Neurology. 1993; 43(1): 61–4.
   PubMed Abstract | Publisher Full Text
- 144. Santos-Lozano A, Villamandos García D, Sanchis-Gomar F, et al.: Niemann-Pick disease treatment: a systematic review of clinical trials. Ann Transl Med. 2015; 3(22): 360.
  - PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- Xu M, Liu K, Swaroop M, et al.: 8-Tocopherol reduces lipid accumulation in Niemann-Pick type C1 and Wolman cholesterol storage disorders. J Biol Chem. 2012; 287(47): 39349–60.
  - PubMed Abstract | Publisher Full Text | Free Full Text
- 146. Liu B, Turley SD, Burns DK, et al.: Reversal of defective lysosomal transport in NPC disease ameliorates liver dysfunction and neurodegeneration in the npc1\* mouse. Proc Natl Acad Sci U S A. 2009; 106(7): 2377–82. PubMed Abstract | Publisher Full Text | Free Full Text
- 147. Liu B, Ramirez CM, Miller AM, et al.: Cyclodextrin overcomes the transport defect in nearly every organ of NPC1 mice leading to excretion of sequestered cholesterol as bile acid. J Lipid Res. 2010; 51(5): 933–44. PubMed Abstract | Publisher Full Text | Free Full Text
- Davidson CD, Ali NF, Micsenyi MC, et al.: Chronic cyclodextrin treatment of murine Niemann-Pick C disease ameliorates neuronal cholesterol and glycosphingolipid storage and disease progression. PLoS One. 2009; 4(9): e6951
  - PubMed Abstract | Publisher Full Text | Free Full Text
- 149. Vite CH, Bagel JH, Swain GP, et al.: Intracisternal cyclodextrin ameliorates neurological dysfunction, increases survival time, and stops Purkinje cell death in feline Niemann-Pick type C1 disease. Mol Genet Metab. 2015; 114(2): S122
  - Publisher Full Text | F1000 Recommendation
- Aqul A, Liu B, Ramirez CM, et al.: Unesterified cholesterol accumulation in late endosomes/lysosomes causes neurodegeneration and is prevented by driving cholesterol export from this compartment. J Neurosci. 2011; 31(25): 9404–13.
   PubMed Abstract | Publisher Full Text | Free Full Text
- 151. Ottinger EA, Kao ML, Carrillo-Carrasco N, et al.: Collaborative development of 2-hydroxypropyl-β-cyclodextrin for the treatment of Niemann-Pick type C1 disease. Curr Top Med Chem. 2014; 14(3): 330–9. PubMed Abstract | Publisher Full Text | Free Full Text
- 152. F Dai S, Dulcey AE, Hu X, et al.: Methyl-β-cyclodextrin restores impaired autophagy flux in Niemann-Pick C1-deficient cells through activation of AMPK. Autophagy. 2017; 13(8): 1435–51.

  PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- 153. Yergey AL, Blank PS, Cologna SM, et al.: Characterization of hydroxypropyl-beta-cyclodextrins used in the treatment of Niemann-Pick Disease type C1. PLoS One. 2017; 12(4): e0175478.
  PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- 154. F Kondo Y, Tokumaru H, Ishitsuka Y, et al.: In vitro evaluation of 2-hydroxyalkylated β-cyclodextrins as potential therapeutic agents for Niemann-Pick Type C disease. Mol Genet Metab. 2016; 118(3): 214–9.
  PubMed Abstract | Publisher Full Text | F1000 Recommendation
- 155. F Ory DS, Ottinger EA, Farhat NY, et al.: Intrathecal 2-hydroxypropyl-β-

- cyclodextrin decreases neurological disease progression in Niemann-Pick disease, type C1: a non-randomised, open-label, phase 1-2 trial. *Lancet.* 2017; **390**(10104): 1758–68.
- PubMed Abstract | Publisher Full Text | F1000 Recommendation
- 156. Pontikis CC, Davidson CD, Walkley SU, et al.: Cyclodextrin alleviates neuronal storage of cholesterol in Niemann-Pick C disease without evidence of detectable blood-brain barrier permeability. J Inherit Metab Dis. 2013; 36(3): 491–8.
  PubMed Abstract | Publisher Full Text | Free Full Text
- 157. Chien YH, Shieh YD, Yang CY, et al.: Lung toxicity of hydroxypropyl-β-cyclodextrin infusion. Mol Genet Metab. 2013; 109(2): 231–2.
  PubMed Abstract | Publisher Full Text
- 158. Crumling MA, Liu L, Thomas PV, et al.: Hearing loss and hair cell death in mice given the cholesterol-chelating agent hydroxypropyl-β-cyclodextrin. PLoS One. 2012; 7(12): e53280.
  PubMed Abstract | Publisher Full Text | Free Full Text
- 159. Kieran D, Kalmar B, Dick JR, et al.: Treatment with arimoclomol, a coinducer of heat shock proteins, delays disease progression in ALS mice. Nat Med. 2004; 10(4): 402-5. PubMed Abstract | Publisher Full Text
- Neef DW, Jaeger AM, Thiele DJ: Heat shock transcription factor 1 as a therapeutic target in neurodegenerative diseases. Nat Rev Drug Discov. 2011; 10(12): 930–44.
   PubMed Abstract | Publisher Full Text | Free Full Text
- Muchowski PJ, Wacker JL: Modulation of neurodegeneration by molecular chaperones. Nat Rev Neurosci. 2005; 6(1): 11–22.
   PubMed Abstract | Publisher Full Text
- 162. Kirkegaard T, Roth AG, Petersen NH, et al.: Hsp70 stabilizes lysosomes and reverts Niemann-Pick disease-associated lysosomal pathology. Nature. 2010; 463(7280): 549–53.
  PubMed Abstract | Publisher Full Text | F1000 Recommendation
- 163. F Kirkegaard T, Gray J, Priestman DA, et al.: Heat shock protein-based therapy as a potential candidate for treating the sphingolipidoses. Sci Transl Med. 2016; 8(355): 355ra118.
  - PubMed Abstract | Publisher Full Text | F1000 Recommendation
- Nylandsted J, Gyrd-Hansen M, Danielewicz A, et al.: Heat shock protein 70 promotes cell survival by inhibiting lysosomal membrane permeabilization. J Exp Med. 2004; 200(4): 425–35.
   PubMed Abstract | Publisher Full Text | Free Full Text
- 165. Nakasone N, Nakamura YS, Higaki K, et al.: Endoplasmic reticulum-associated degradation of Niemann-Pick C1: evidence for the role of heat shock proteins and identification of lysine residues that accept ubiquitin. J Biol Chem. 2014; 289(28): 19714–25.
  PubMed Abstract | Publisher Full Text | Free Full Text
- 166. Ingemann L, Kirkegaard T: Lysosomal storage diseases and the heat shock response: convergences and therapeutic opportunities. J Lipid Res. 2014; 55(11): 2198–210. PubMed Abstract | Publisher Full Text | Free Full Text
- Lanka V, Wieland S, Barber J, et al.: Arimoclomol: a potential therapy under development for ALS. Expert Opin Investig Drugs. 2009; 18(12): 1907–18.
   PubMed Abstract | Publisher Full Text
- Cudkowicz ME, Shefner JM, Simpson E, et al.: Arimoclomol at dosages up to 300 mg/day is well tolerated and safe in amyotrophic lateral sclerosis. Muscle Nerve. 2008; 38(1): 837–44.
   PubMed Abstract | Publisher Full Text
- 169. F Alam MS, Getz M, Haldar K: Chronic administration of an HDAC inhibitor treats both neurological and systemic Niemann-Pick type C disease in a mouse model. Sci Transl Med. 2016; 8(326): 326ra23.
  PubMed Abstract | Publisher Full Text | F1000 Recommendation
- Helquist P, Maxfield FR, Wiech NL, et al.: Treatment of Niemann--pick type C disease by histone deacetylase inhibitors. Neurotherapeutics. 2013; 10(4): 688–97.
  - PubMed Abstract | Publisher Full Text | Free Full Text
- 171. Pipalia NH, Cosner CC, Huang A, et al.: Histone deacetylase inhibitor treatment dramatically reduces cholesterol accumulation in Niemann-Pick type C1 mutant human fibroblasts. Proc Natl Acad Sci U S A. 2011; 108(14): 5620–5. PubMed Abstract | Publisher Full Text | Free Full Text
- 172. Kim SJ, Lee BH, Lee YS, et al.: Defective cholesterol traffic and neuronal differentiation in neural stem cells of Niemann-Pick type C disease improved by valproic acid, a histone deacetylase inhibitor. Biochem Biophys Res Commun. 2007; 360(3): 593–9.
  PubMed Abstract | Publisher Full Text
- 173. Munkacsi AB, Chen FW, Brinkman MA, et al.: An "exacerbate-reverse" strategy in yeast identifies histone deacetylase inhibition as a correction for cholesterol and sphingolipid transport defects in human Niemann-Pick type C disease. J Biol Chem. 2011; 286(27): 23842–51. PubMed Abstract | Publisher Full Text | Free Full Text
- 174. Di XJ, Han DY, Wang YJ, et al.: SAHA enhances Proteostasis of epilepsy-associated a1(A322D)β2/2 GABA, receptors. Chem Biol. 2013; 20(12): 1456–68. PubMed Abstract | Publisher Full Text | Free Full Text
- 175. Calamini B, Morimoto RI: Protein homeostasis as a therapeutic target for

- diseases of protein conformation. Curr Top Med Chem. 2013; 12(22): 2623–40. PubMed Abstract | Publisher Full Text | Free Full Text
- 176. Yang C, Rahimpour S, Lu J, et al.: Histone deacetylase inhibitors increase glucocerebrosidase activity in Gaucher disease by modulation of molecular chaperones. Proc Natl Acad Sci U S A. 2013; 110(3): 966–71. PubMed Abstract | Publisher Full Text | Free Full Text
- 177. Newton J, Hait NC, Maceyka M, et al.: FTY720/fingolimod increases NPC1 and NPC2 expression and reduces cholesterol and sphingolipid accumulation in Niemann-Pick type C mutant fibroblasts. FASEB J. 2017; 31(4): 1719–30. PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- 178. Verot L, Chikh K, Freydière E, et al.: Niemann-Pick C disease: functional characterization of three NPC2 mutations and clinical and molecular update on patients with NPC2. Clin Genet. 2007; 71(4): 320–30. PubMed Abstract | Publisher Full Text
- 179. Bonney DK, O'Meara A, Shabani A, et al.: Successful allogeneic bone marrow

- transplant for Niemann-Pick disease type C2 is likely to be associated with a severe 'graft versus substrate' effect. *J Inherit Metab Dis.* 2010; **33**(Suppl 3): S171–3
- PubMed Abstract | Publisher Full Text
- 180. Xie C, Gong XM, Luo J, et al.: AAV9-NPC1 significantly ameliorates Purkinje cell death and behavioral abnormalities in mouse NPC disease. J Lipid Res. 2017; 58(3): 512–8.

  PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
- 181. Chandler RJ, Williams IM, Gibson AL, et al.: Systemic AAV9 gene therapy improves the lifespan of mice with Niemann-Pick disease, type C1. Hum Mol Genet. 2017; 26(1): 52–64.
  PubMed Abstract | Publisher Full Text | F1000 Recommendation
- 182. F Di Lazzaro V, Marano M, Florio L, et al.: Niemann-Pick type C: focus on the adolescent/adult onset form. Int J Neurosci. 2016; 126(11): 963–71. PubMed Abstract | Publisher Full Text | F1000 Recommendation

# **Open Peer Review**

<b>Current Referee Status:</b>	<b>~</b>	•	<b>/</b>
--------------------------------	----------	---	----------

## **Editorial Note on the Review Process**

F1000 Faculty Reviews are commissioned from members of the prestigious F1000 Faculty and are edited as a service to readers. In order to make these reviews as comprehensive and accessible as possible, the referees provide input before publication and only the final, revised version is published. The referees who approved the final version are listed with their names and affiliations but without their reports on earlier versions (any comments will already have been addressed in the published version).

## The referees who approved this article are:

### Version 1

- Maria D Ledesma Centro Biología Mole, CSIC-UAM, Madrid, Spain Competing Interests: No competing interests were disclosed.
- Heiko Runz Merck & Co., Kenilworth, NJ, USA Competing Interests: HR is an employee to Merck & Co. He is or has been scientific advisor to Actelion, Vtesse and Sucampo Inc.
- Andrew P Lieberman Department of Pathology, University of Michigan Medical School, Ann Arbor, MI, USA Competing Interests: No competing interests were disclosed.

The benefits of publishing with F1000Research:

- Your article is published within days, with no editorial bias
- You can publish traditional articles, null/negative results, case reports, data notes and more
- The peer review process is transparent and collaborative
- Your article is indexed in PubMed after passing peer review
- Dedicated customer support at every stage

For pre-submission enquiries, contact research@f1000.com

